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(57) Abstract

Provided herein is a pharmaceutical composition containing one or more DNA molecules encoding fragments of a protoin overexpressed in tumor cells, in order to induce an anti-tumor Ag-specific immune response, in association with suitable excipients and adjuvants.

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PHARMACEUTICAL COMPOSITION, CONTAINING FRAGMENTS OF AN ANTIGENIC
PROTETN ENCODING DNA ENDOWED WITH ANTI-TIMOR EFFECT.

Field of the invention

The invention relates to a pool of DNA plasmid constructs containing the sequences of human MUC-1 encoding fragments and to a pool of DNA plasmids in which the fragments themselves are preceded by the sequence encoding a protein consisting of human ubiquitin fused to a bacterial LacI fragment. The invention further relates to their use in the preparation of pharmaceutical compositions for use as DNA anti-tumor vaccines.

Background art

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The invention provides an anti-tumor therapy based on the induction or activation of the immune response able to bring about tumor rejection. The validity of such an idea is demonstrated from the first clinical results; for example, patients treated with a vixal vaccine containing the Carcinoembryonic Antigen (CEA) encoding sequences demonstrated immune system activation against this antigen (Tsang KY et al. J. Natl. Cancer. Inst. 87: 982, 1995).

The activation of an immune anti-tumor response is achievable through four different approaches:

 a) Ex vivo engineering of patient tumor cells in order to make them more immunoquaic and suitable as a vaccine; | 10mm | 10mm

- b) Ex vivo engineering of patient immune cells in order to pre-activate an in vitro immune response.
- c) Inoculation of naked or liposome capsulated or viral particle integrated (retrovirus, vaccinia virus, adenovirus, etc.) DNA encoding tumor associated antigens;
- d) Treatment with recombinant or synthetic soluble tumor antigens conjugated or mixed with adjuvants.

The first two approaches consist of the engineering of every single patient cell and are limited in that they are necessarily patient-specific, while the latter two are aimed to

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obtain products comparable to a traditional drug.

The new vaccination methods reflect the development of new technologies. The recent indications coming from experimentation on DNA naked vaccines that induce either a persistent antibody or a cell immune response, make the traditional protein subunit vaccines constituted of certain specific peptides, inducing a lymphocyte population, obsolete. Intramuscularly or intradermically injected proteins, encoded by naked DNA, induce a cytotoxic-specific response as well as a helper response. This powerful combination is extremely effective but the underling mechanism is not completely clarified yet. Muscle cells express class I MHC antigens at low levels only, and do not apparently express class II antigens or co-stimulatory molecules. Consequently, transfected muscle cells are unlikely to play an important role in the onset of the immune response per se. Recent data show that Antigen Presenting Cells (APC), such as macrophages or dendritic cells, play a fundamental role in capturing the myocyte released antigen and in the subsequent processing and presenting of the respective peptides in the context of the class I and II molecules, thus inducing a CD8+ cell activation with cytotoxic activity as well as activation of the CD4+ cells co-operating with B lymphocytes in eliciting the antibody response (Corr M et al J. Exp. Med. 184:1555, 1996) (Tighe, H. et al. Immunology Today 19:89, 1998).

... Furthermore, the use of cytokines is known to improve the therapeutic effect deriving from immunization with DNA. Cytokines can be administered in the form of exogenous proteins as reported in Irvine et al., J. Immunol. 156: 238, 1996. An alternative approach is represented by the contemporaneous inoculation of both the tumor antigen or the desired cytokine encoding plasmids, thus allowing the cytokine to be produced in situ (Kim JJ et al. Immunol 158: 816, 1997).

The active immunization approach of the present invention is based on the use of DNA vectors as vaccines against the MUC-1

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antigen orPolymorphic Epithelial Mucin overexpressed in tumor cells. MUC-1 is an epithelial luminal surface glycoprotein (Patton S. et al. BBA 1241:407, 1995). In the cell transformation process this glycoprotein loses the apical localization and its expression level rises dramatically. The protein function consists of protecting the luminal surfaces, for example in the mammal gland, ovary, endometrium, colon, stomach, pancreas, bladder, kidney, etc. A glycosylation defect is reported that makes tumor cell associated MUC-1 antigenically different from normal cell associated MUC-1. This phenomenon causes tumor MUC-1 to expose the antigen epitopes that are normally masked by the sugar moieties in the normal cell expressed MUC-1. This characteristic makes tumor MUC-1 particularly interesting in an induction of a tumor specific antibody response (Apostolopoulos V. et al. Crit. Rev. Immunol. 14:293, 1994).

As an objective, the vaccination is aimed at inducing immune responses against tumor cells expressing MUC1 at high levels, preserving at the same time the low expressing normal epithelia. The DNA vaccination relies upon the entrance of a gene or portions thereof inside the body cells followed by transcription and translation of the inserted sequence and thus the intracellular synthesis of the corresponding polypeptide. An important advantage of this system is that the neo-synthesized protein is naturally processed inside the cell and the produced peptides are associated with the Major Histocompatibility Complex class I molecules (MHC-I). The MHC/peptide complexes are therefore naturally exported to the cell surface where they can be recognized by the immune system CD8+ cytotoxic cells. Only the polypeptides synthesized inside the cell are then processed and presented in association with the MHC class I molecules, thus making it the only mechanism to stimulate, a specific cytotoxic response. Vaccination systems based on protein or peptide administration are usually more effective in stimulating

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the antibody immune response which, to date, has been shown to be ineffective in rejecting tumor cells. Current gene therapy techniques rely upon DNA packaging in recombinant viral vectors (retrovirus and adenovirus). The naked DNA administration is much more advantageous in terms of effectiveness and safety compared to viral vector therapies (Kumar V and Sercarz E. Nature Med. 2: 857, 1996; McDonnel NM et al., New England J. of Med. 334: 42, 1996). In fact naked DNA is unable either to duplicate or integrate in the host tissue DNA and does not induce the immune response to viral proteins.

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The use of the ubiquitin to enhance the neo-synthesized protein processing and thus cytotoxic lymphocyte induction was recently reported (Rodriguez F. et al., J. Virology 71: 8497, 1997). The use of ubiquitin in order to generate proteins with an N-terminal amino acid, usaking them unstable and thus prone to enhanced degradation, had been previously reported (Bechmair A. et al., SCIENCE 234: 179, 1986). The higher instability of these proteins was subsequently related to enhanced intracellular processing and presentation of model proteins by MHC-1 (Grant B P et al., J. Immunol. 155: 3750, 1995) (Mu Y and Kipps T.J., J. Immunol. 159: 6037, 1997).

The use of single constructs containing partial antigen encoding DNA fragments (influenza virus nucleoprotein), having a higher antigenic presentation efficiency compared to the analogues with the whole antigenic sequence, in DNA vaccination was reported (Anton L. C. et al., J. Immunol. 158: 2535, 1997). Furthermore the processing of intracellular proteins and presentation of the respective peptides by MNC class I proteins physiologic conditions, in underlie the mechanism immunological surveillance. For a given protein and a specific MHC context, there are peptide fragments termed dominants (i. e. prevailing on subdominants or cryptics), which are unable to generate any immune response because they are recognized as "self". It has now been outlined, according to an aspect of the

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present invention, that an approach aimed at supporting the nondominant epitope presentation by the administration of a mix of antigen protein fragments is able to elicit a surprising cytotoxic immune response.

Description of the invention

It has now been found that DNA molecules, encoding fragments of a protein overexpressed in tumor cells, can be conveniently used to induce an antigen-specific anti-tumor immune response.

The invention relates particularly to a pharmaceutical composition containing one or more DNA encoding Mucin (MUC-1) protein fragments.

The DNA used in the present invention can be plasmid or viral DNA, preferably plasmid DNA obtained employing the pMRS30 expression vector described in fig. 13.

The compositions according to the invention contain preferably at least two DNA fragments of the Mucin (MUC-1) or of another protein overexpressed in tumor cells.

The compositions according to the invention contain preferably at least four fragments, each ranging from 200 to about 700 nucleotides, each sequence being juxtaposed and possibly partially overlapping, from about 50 to about 150 nucleotides, at the 3' and/or 5' end of the adjacent one.

The DNA fragments according to the invention can be possibly preceded at the 5' end by a ubiquitin encoding DNA sequence and possibly also by a LacI portion of Escherichia coli.

The invention relates also to new DNA fragments and to the use of Mucin-1 fragments defined above in the medicine and antitumor vaccine preparation.

Description of the figures

Fig. 1

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the Xbal site of the pMRS166 expression vector. This DNA includes the sequence corresponding to mucleotides 136-339 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by the two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the amino acids encoded by the 136-339 fragment of the EMBL sequence J05581.

Fig. 2

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Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS169 expression vector. This DNA includes the sequence corresponding to nucleotides 205-720 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the amino acids encoded by the 205-720 fragment of the EMBL sequence J05581.

Fig. 3

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS168 expression vector. This DNA includes the sequence corresponding to nucleotides 631-1275 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the amino acids encoded by the 631-1275 fragment of the EMBL sequence J05581.

Fig. 4

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS167 expression vector. This DNA includes the sequence corresponding to nucleotides 1222-1497 of the EMBL sequence J05581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the

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amino acids encoded by the 1222-1497 fragment of the EMBL sequence J05581.

Fig. 5

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS175 expression vector. This DNA includes the sequence corresponding to nucleotides 136-1497 of the EMBL sequence JO5581, preceded by the translation start codon, ATG and followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes a Metionin followed by the amino acids encoded by the 136-1497 fragment of the EMBL sequence JO5581.

Fig. 6

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Nucleotide DNA sequence (with the respective amino acid sequence) termed UBILacI. The encoded polypeptide includes the Ubiquitin sequence fused to a partial sequence of the bacterial protein beta-galactosidase, as described in Chau V. et al. Science 243: 1576, 1989.

Fig. 7

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the expression vector pMRS30 to give the pMRS171 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence corresponding to nucleotides 136-339 of the EMBL sequence J05581 followed by two translation stop codons, TGA and TAA. The coded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 136-339 of the EMBL sequence J05581.

Fig. 8

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS174 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 205-720 of the EMBL

sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 205-720 of the EMEL sequence J05581.

Fig. 9

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Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS173 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 631-1275 of the EMBL sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 631-1275 of the EMBL sequence J05581.

Fig. 10

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS172 expression vector. This DNA includes the sequence termed UBILacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 1222-1497 of the EMBL sequence J05581 followed by two translation stop codons, TGA and TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 1222-1497 of the EMBL sequence J05581.

Fig. 11

Nucleotide DNA sequence (with the respective amino acid sequence) inserted at the XbaI site of the pMRS30 expression vector to give the pMRS176 expression vector. This DNA includes the sequence named UBLLacI (see fig. 6) fused to the sequence partially corresponding to nucleotides 136-1497 of the EMBL sequence J05581 followed by two translation stop codons, TGA and

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TAA. The encoded polypeptide thus includes the amino acid sequence reported in Fig. 6, fused to the sequence including the amino acids encoded by the fragment 136-1497 of the EMBL sequence J05581.

Fig. 12

Electrophoretic analysis on 1% agarose gel in 1X TBE. mRNA extracted from CHO, CD34+ dendritic cells and dendritic cells from PBMC, respectively, transfected with pMRS169, and subjected to RT-PCR reaction either with (lanes 4, 8, 12) or without (lanes 5, 9, 13) Reverse Transcriptase. Molecular weight DNA marker (lane 1); internal negative controls (lanes 2, 6); internal positive controls (lanes 3, 7, 10, 11); positive control from Promeaa kit (lane 14).

Fig. 13

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Nuclectide sequence of the pMRS30 expression vector. The 1-2862 region corresponds to the AccI (location 504) - BamHI (location 3369) region of the pSV2CAT vector (BMBL M77788); the 2863-3721 region includes the human cytomegalovirus promoter (human cytomegalovirus major immediate-early gene enhancer); the 3722-4905 region includes several cloning sites, including XbaI (location 3727), and the processing signal of the rabbit betaclobin cene.

Detailed description of the invention

A DNA plasmid pool encoding, in eukaryotic cells, fragments of the MUC-1 human protein antigen was prepared. Constructs are based on the mammalian expression vector termed pMRS30, described in figure 13 and previously claimed in the Patent Application W095/11982, and contain partial sequences of the MUC-1 cDNAs reported in the EMBL database with accession number J05581. MUC-1 encoding DNA was fragmented so that each fragment represents a discrete portion, partially overlapping to the adjacent ones. Administration of a mix of such plasmids can cause different plasmids to transfect different APC cells at the administration site. Therefore such cells produce and process

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The present invention thus relates to the use of a group of four constructs (Figures 1 to 4) containing MUC-1 cDNA partial fragments in admixture containing at least two of them and a group of four constructs (Figures 7 to 10) containing MUC-1 cDNA partial fragment preceded by the DNA encoding a protein sequence containing Ubiquitin and an Escherichia coli Lac I portion (Figure 6) used separately or in admixture containing at least two of them.

The present invention relates also to the use of the construct (Figure 5) containing the almost complete sequence of the MUC-1 cDNA and the construct (Figure 11) containing the almost complete sequence of the MUC-1 cDNA preceded by the DNA encoding a protein sequence containing Ubiquitin and an Escherichia coli Lec I portion.

The mixture of the four constructs containing the partial fragments of the MUC-1 cDNA and the mixture of the four constructs containing the partial fragments of the MUC-1 cDNA preceded by the DNA encoding a protein sequence, containing Ubiquitin and an Escherichia coli Lac I portion, represents a preferred embodiment of the present invention.

Constructs according to the present invention can be used in the anti-tumor therapy of patient affected with tumors characterized by high MUC-1 expression.

Constructs described in the present invention were obtained as follows.

In the case of the first series of constructs, the fragments of the MUC-1 DNA were obtained by RT-PCR from BT20 cell line or by DNA partial chemical synthesis. Such fragments were then cloned into the pMRS30 expression vector and verified by sequencing.

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In the case of the second series of constructs, the fragments were obtained from the first series of constructs by a PCR re-amplification. These fragments were then fused to the DNA encoding the Ubiquitin (obtained by RT-PCR from MCF7 cell line mRNA) and a partial lacI sequence (obtained by PCR from the commercial vector pGEX). DNA sequences thus obtained were then cloned in the pMRS30 expression vector and verified by sequencing. For the intended therapeutic or prophylactic uses, fragments or constructs according to the invention are suitably formulated, using carriers and methods previously employed in naked DNA vaccines, as described for example in The Immunologist, 1994, 2:1; WO 90/11092, Proc. Natl. Acad. Sci. U.S.A., 1986, 83, 9551; US 5580859; Immunology today 19 (1998). 89-97); Proc. Natl. Acad. Sci. U.S.A. 90 (1993), 11478-11482; Nat. Med. 3 (1997), 526-532; Vaccine 12 (1994), 1495-1498; DNA Cell. Biol. 12 (1993), 777-783. The dosages will be determined on the basis of clinical and pharmacological-toxicological trials. Generally speaking, they will be comprised between 0.005 ug/kg and 5 ug/kg of the fragment mix. The composition of the invention can also contain a cytokine or a cytokine encoding plasmid.

The invention will be further illustrated by means of the following examples.

Example 1. Plasmid pMRS166 construction.

BT20 tumor cells (ATCC HTB-19) were cultured in Eagles MRM supplemented with 10% fetal calf serum. Ten million cells were trypsinized, washed with PBS, and mRNA extracted.

An aliquot of this RNA was subjected to RT-PCR (reverse transcriptase-polymerase chain reaction) reaction in the presence of the following synthetic oligonucleotides:

V11 (5 GATCTCTAGAATGACAGGTTCTGGTCATGCAAGC 3)

V4 (5 GATCTCTAGAAAGCTTATCAACCTGAAGCTGGTTCCGTGGC 3)

The produced DNA fragment, purified and digested with the restriction enzyme XbaI, was cloned into the bMRS30 expression

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vector, containing the human cytomegalovirus promoter and the beta-globin polyadenylation signal as claimed in the Patent W09511982. The resulting pMRS166 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the nucleotides 136-339 of the EMBL sequence J05581, and two stop codons. TGA and TAA.

This fragment is reported in fig. 1.

Example 2. Plasmid pMRS169 construction.

An aliquot of the RNA obtained as reported in example 1 was amplified by RT-PCR in the presence of the following synthetic oligonuclotides:

V12 (5 GATCTCTAGAATGGTGCCCAGCTCTACTGAGAAGAATGC 3)

V15 (5 GGCGGTGGAGCCCGGGGCTGGCTTGT 3)

The produced DNA fragment, purified and digested with the restriction enzymes SmaI and XbaI, was fused, by the SmaI restriction site, to a DNA fragment entirely synthetically constructed, and including a sequence partially corresponding to the muleotides 457-720 of the EMBL sequence J05581 and two stop codons, TGA and TAA. The whole fragment was thus cloned in the XbaI site of the pMRS30 expression vector. The resulting pMRS169 vector contains a DNA fragment including the ATG codon, the sequence partially corresponding to the nucleotides 205-720 of the EMBL sequence J05581, and two stop codons, TGA and TAA.

This fragment is reported in fig. 2.

Example 3. Plasmid pMRS168 construction.

An aliquot of the RNA obtained as reported in example 1 was amplified by RT-PCR in the presence of the following synthetic oligonuclotides:

V13 (5 GATCTCTAGAATGGGCTCAGCTTCTACTCTGGTGCACAACGGC 3)

V8 (5 GATCTCTAGAAAGCTTATCACAAGGCAATGAGATAGACAATGGCC 3)

The produced DNA fragment, purified and digested with the restriction enzyme XbaI was cloned in the pMR830 expression vector. The resulting pMR8168 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the

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nucleotides 631-1275 of the EMBL sequence J05581, and two stop codons, TGA and TAA.

This fragment is reported in fig. 3.

Example 4. Plasmid pMRS167 construction.

An aliquot of the RNA obtained as reported in example 1 was subjected to RT-PCR reaction in the presence of the following synthetic oligonucleotides:

V14 (5 GATCTCTAGAATGCTGGTGCTGGTCTGTGTTCTGGTTGCGC 3)

V10 (5 GATCTCTAGAAAGCTTATCACAAGTTGGCAGAAGTGGCTGC 3)

The produced DNA fragment, purified and digested with the restriction enzyme XbaI was cloned in the pNRS30 expression vector. The resulting pNRS167 vector contains a DNA fragment including the ATG codon, the sequence corresponding to the nucleotides 1222-1497 of the EMBL sequence JO5581, and two stop codons. TGA and TAA.

This fragment is reported in fig. 4.

Example 5. Plasmid pMRS175 construction.

pMRS166, 169, 168, 167 plasmids were subjected to PCR reaction in the presence of the following nucleotide pairs:

V11 (see example 1)

V18 (5 AACCTGAAGCTGGTTCCGTGGC 3) for pMRS166

V19 (5 GTGCCCAGCTCTACTGAGAAGAATGC 3)

V20 (5 GCTGGGAATTGAGAATGGAGTGCTCTTGC 3) for pMRS169

V21 (5 GGCTCAGCTTCTACTCTGGTGCACAACGGC 3)

V22 (5 CAAGGCAATGAGATAGACAATGGCC 3) for pMRS168

V23 (5 CTGGTGCTGGTCTGTGTTCTGGTTGCG 3)

V10 (see example 4) for pMRS167

The four DNA fragments obtained in the respective PCR reactions were mixed in equimolar amounts and PCR reacted in the presence of the V11 and V10 oligonuclotides.

The produced DNA fragment, purified and digested with the XDaI restriction enzyme, was cloned in the pMRS30 expression vector. The resulting pMRS175 vector contains a DNA fragment including the ATG codon, the sequence partially corresponding to

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the nucleotides 136-1497 of the EMBL sequence J05581 and two stop codons TGA and TAA:

This fragment is reported in fig. 5.

Example 6. Plasmid pMRS171 construction.

MCP7 tumor cells (ATCC HTB-22) were cultured in Eagles MEM supplemented with 10% fetal calf serum. Ten million cells were trypsinized, washed with PBS, and mRNA extracted.

An aliquot of this RNA was subjected to RT-PCR in the presence of the following synthetic oligonucleotides:

UBlup (5GATCTCTAGAATGCAGATCTTCGTGAAGACCCTGACTGGT 3)
UBldown

(5TCACCAGCGAGACGGGCAACAGCCATGCACCACTACCGTGCCTCCCACCTCTGAGACGGAGC ACCAGG 3)

The reaction produces a DNA fragment termed fragment 1.

DNA from pGEX11T (Pharmacia) was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

LacIup (5CCTCCGTCTCAGAGGTGGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTGAAAAG 3)

Lacidown (5GATCGGATCCTCGGGAAACCTGTCGTGCCAGCTGC 3)

This reaction gives a DNA fragment termed fragment 2.

The 1 and 2 DNA fragments, obtained in the respective FCR reactions, were mixed in equimolar amounts and subjected to PCR reaction in presence of the UBIup and LacIdown oligonucleotides.

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was cloned into the pUCLS commercial plasmid. The resulting pMRS156 vector contains a DNA fragment including the sequence encoding the ubiquitin fused to the sequence encoding a bacterial beta-galactosidase portion. This fragment, termed UBILacI, is reported in fig. 6.

Plasmid pMRS166 DNA was subjected to a PCR reaction in presence of the following synthetic oligonucleotides:

V3 (5GATCGGATCCACAGGTTCTGGTCATGCAAGC 3)

V4 (see Example 1)

The produced DNA fragment, purified and digested with the

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restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS171 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 136-339 nucleotides of the EMBL sequence J05581 and two stop codons, TGA and TAA. This fragment is reported in fig. 7.

Example 7. Plasmid pMRS174 construction.

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Plasmid pMRS169 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

V6 (5GATCTCTAGAAAGCTTATCAGCTGGGAATTGAGAATGGAGTGCTCTTGC 3)

V5 (5GATCGGATCCGTGCCCAGCTCTACTGAGAAGAATGC 3)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS310 expression vector. The resulting pMRS174 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 205-720 nucleotides of the EMBL sequence J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 8.

Example 8. Plasmid pMRS173 construction.

Plasmid pMRS168 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

V7 (5GATCGGATCCGGCTCAGCTTCTACTCTGGTGCACAACGGC 3)

V8 (see example 3)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from the pMRS156 plasmid. The resulting fragment was cloned into the pMRS306 expression vector. The resulting pMRS173 vector contains a DNA fragment including the UBILscI sequence, the sequence corresponding to the 631-1275 nucleotides of the EMBL sequence

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J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 9.

Example 9, Plasmid pMRS172 construction.

Plasmid pMRS167 DNA was subjected to PCR reaction in the presence of the following synthetic oligonucleotides:

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V9 (5 GATCGGATCCCTGGTGCTGGTCTGTGTTCTGGTTGCGC 3)

V10 (see example 4)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from PMRS156 plasmid. The resulting fragment was cloned into the PMRS30 expression vector. The resulting pMRS172 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 1222-1497 nucleotides of the EMBL sequence J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 10.

Example 10. Plasmid pMRS176 construction.

Plasmid pMRS167 DNA was subjected PCR reaction in the presence of the following synthetic oligonucleotides:

V3 (see example 6)

V10 (see example 4)

The produced DNA fragment, purified and digested with the restriction enzymes XbaI and BamHI, was fused, by ligation into the two BamHI sites, to the UBILacI fragment deriving from pMRS156 plasmid. The resulting fragment was cloned into the pMRS30 expression vector. The resulting pMRS176 vector contains a DNA fragment including the UBILacI sequence, the sequence corresponding to the 136-1497 nucleotides of the EMBL sequence J05581, and two stop codons, TGA and TAA. This fragment is reported in fig. 11.

Example 11. Eukaryotic cell transfection and testing for transcription.

CHO (Chinese Hamster Ovary) cells were cultured in alpha MEM supplemented with ribonucleotides and deoxyribonucleotides

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at transfection time.

Dendritic cells were obtained from CD34+ hemopoietic precursors cultured in IMDM without serum, supplemented with GM-CSF, IL4, SCF, Flt3 and INFalpha. After 7 days the obtained cell population was transfected.

Dendritic cells were obtained from monocytes isolated from PEMC (peripheral blood mononuclear cells), cultured in RPMI supplemented with FCS, GM-CSF, and IL-4. After 7 days the obtained cell population was transfected.

In each case, about one million cells were transfected with one of the plasmids reported in examples 1 to 10. Transfection was carried out using 3 µg of plasmid DNA and 4 µl of DMRIE (Gibco) by lipofection.

After 24 hours cells were harvested, washed with PBS and lysed in order to extract the mRNA.

A mRNA aliquot was subjected to RT-PCR reaction in the presence of the oligonucleotide pair specific for the transfected DNA plasmid.

This experiment was carried out for each plasmid reported in the examples 1 to 10, using the following oligonucleotide pairs: V11/V4 for pMRS166, V12/V6 for pMRS169, V13/V8 for pMRS168, V4/V10 for pMRS167, V4/V10 for pMRS175, UBlup/V4 for pMRS171, UBlup/V6 for pMRS174, UBlup/V8 for pMRS173, UBlup/V10 for pMRS172, V14/V10 for pMRS176.

As a representative example, figure 12 reports the electrophoretic analysis of the DNA fragments obtained by RT-PCR from the mRNA of the three cell populations, transfected with the pNRS169 plasmid. In this case the oligonucleotide pair V12/V6 was used.

Example 12. In vivo study results.

In the in vivo studies, the mixtures of the four fragments and the pMRS30 plasmid (vector without insert and thus used as a negative control) were used. In order to test the occurred immunization, an ELISA test was used to show the human mucin

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specific antigens.

The in vivo studies were conducted using human MUC1 transgenic C57BL mice. As a consequence in these animals the MUC1 protein represents a self-protein. The employed vaccination schedule consists of 3 intradermic (dorsal portion, 50 micrograms DNA for each side) administrations (at days 0, 14, 28) of 100 micrograms plasmid DNA. At day 14 after the last administration, the animals were sacrificed and sera were tested for anti-human mucin antibodies.

The assayed fragment mixes, object of the present invention, stimulated a good immune response in the treated animals.

On the other hand, vaccination experiments with a 60-aminoacid peptide corresponding to the 20 aminoacids reported in fig. 2, from location 86 to location 105, repeated three times (this peptide is termed 3XTR), were also carried out.

The two vaccinations differ in the type of the elicited antibody response. The antibody titer results much more higher in the vaccination with 3XTR. Furthermore the noticed IgG subtypes are in favor of an essentially humoral (antibody) response in the case of vaccination with 3XTR, and of a cellular response (cytotoxic) in the case of vaccination with DNA. For anti-tumor therapy, a principally cytotoxic immune response is preferable. Because the experiments were carried out on transgenic mice, in whom the human mucin is "self", we can foresee a similar response in humans. This response could justify the use, as DNA vaccines, of the compounds of the present invention in the treatment of MUC1 overxpressing human tumors.

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- 1. Pharmaceutical composition containing one or more DNA molecules, encoding fragments of a protein overexpressed in tumor cells in order to induce an antitumor Ag-specific immune response, in combination with suitable excipients and adjuvants.
- Pharmaceutical composition according to claim 1 wherein the overexpressed protein is MUC-1.
- 3. Pharmaceutical composition according to claim 1 or 2 containing at least two DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
- Composition according to claim 3 containing at least three DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
- 5. Composition according to claim 4 containing at least four DNA molecules each containing a cDNA sequence encoding a Mucin fragment (MUC-1).
 - 6. Composition according to claims 3, 4 or 5 wherein the DNA sequences comprise about 200 to about 700 nucleotides, each sequence being contiguous and possibly partially overlapping, from about 50 to about 150 nucleotides at the 3' and/or 5' end, to the adjacent one.
 - 7. Pharmaceutical composition according to any claim from 2 to 6 wherein the used mixture consists of, at least, two plasmid DNA molecules, each containing a DNA fragment selected from those whose sequences are described in figures 1, 2, 3, and 4.
 - 8. Pharmaceutical composition according to claim 7 wherein the used mixture consists of the pool of plasmid DNA molecules, where each molecule contains a DNA fragment selected from those whose sequences are described in figures 1, 2, 3, and 4.
 - Pharmaceutical composition according to claim 1 or 2 wherein a plasmid DNA molecule containing the sequence described in figure 5 is used.
 - 10. Pharmaceutical composition according to claims 7, 8, or 9

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wherein the used plasmid DNA molecules derive from the fusion of the pMRS30 expression vector in Fig. 13 to each sequence described in figures 1, 2, 3, 4, 5.

- 11. Pharmaceutical composition according to claims 2 to 6 wherein the used sequences, corresponding to single fragments of the protein, are preceded in the 5' termini by the sequence described in Fig. 6 encoding the ubiquitin and a LacI portion from Escherichia Coli.
- 12. Pharmaceutical composition according to claim 11 wherein the mixture consists of one or more sequences deriving from joining the pMRS30 expression vector, described in Fig. 13, to a DNA sequence selected from those described in figures 7, 8, 9, and 10.
- 13. Pharmaceutical composition according to claim 11 wherein the mixture consists of the totality of the sequences deriving from joining the pMRS30 expression vector to a DNA sequence selected from those described in figures 7, 8, 9, and 10.
 - 14. Pharmaceutical composition according to claim 11 wherein the mixture consists of a sequence deriving from joining the pMRS30 expression vector to the sequence described in figure 11.
 - 15. Pharmaceutical composition according to any preceding claims, further containing a cytokine or a cytokine encoding plasmid.
 - 16. A plasmid DNA molecule consisting of the pMRS30 expression vector joined to a DNA sequence, encoding a MUC-1 protein fragment and whose sequence is selected from the group of those described in figures 1, 2, 3, 4, and 5.
 - 17. A DNA molecule encoding a protein MUC-1 fragment preceded in its 5' terminus by the sequence described in Fig. 6.
- 30 18. A DNA molecule according to claim 17 selected from those described in figures 7, 8, 9, 10, and 11.
 - 19. A plasmid DNA molecule obtained by joining the pMRS expression vector to a DNA molecule selected from those of claim 17 or 18.

20. Use of DNA molecules of claims 16-19 in the preparation of a composition with anti-tumor effect.

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Figure 1

- 1 ATGACAGGTTCTGGTCATGCAAGCTCTACCCCAGGTGGAGAAAAG
- 1 Met Thr GlySer GlyHis At a Ser Ser Thr ProGlyGlyGlyLys
- 46 GAGACTTCGGCTACCCAGAGAAGTTCAGTGCCCAGCTCTACTGAG
- 16 GluThr Ser AlaThr GlnArgSer Ser Val ProSer Ser Thr Glu
- 91 AAGAATGCTGTGAGTATGACCAGCAGCGTACTCTCCAGCCACAGC
- 31 Lys AsnAl a Val Ser Met Thr Ser Ser Val Leu Ser Ser His Ser
- 136 CCCGGTTCAGGCTCCTCCACCACTCAGGGACAGGATGTCACTCTG 46 ProGlySer GlySer Ser Thr Thr GlnGlyGlnAspVal Thr Leu
- 181 GCCCCGGCCACGGAACCAGCTTCAGGTTGATAA
 - 61 A I a Pro Al a Thr Glu Pro Al a Ser Gl v

Figure 2

1 ATCGTGCCCAGCTCTACTGAGAAGAATGCTGTGAGTATGACCAGC 1 Met Val ProSer Ser Thr GluLvs AsnAl aVal Ser Met Thr Ser 46 AGCGTACTCTCCAGCCACAGCCCCGGTTCAGGCTCCTCCACCACT 16 For Val LeuSer Ser His Ser ProGlySer GlySer Ser Thr Thr 91 CAGGGACAGGATGTCACTCTGGCCCCGGCCACGGAACCAGCTTCA 31 FinGlyGlnAspValThrLeuAlaProAlaThrGluProAlaSer 136 GGTTCAGCTGCCACCTGGGGACAGGATGTCACCTCGGTCCCAGTC 46 GiySer AlaAlaThr TrpGiyGlnAspValThr Ser ValProVal 181 ACCAGGCCAGCCTGGGCTCCACCACCCCGCCAGCCCACGATGTC 61 Thr ArgProAl aLeuGlySer Thr Thr ProProAl aHi sAspVal 226 ACCTCAGCCCGGACAACAAGCCAGCCCCGGGAAGTACTGCTCCA 76 ► Thr Ser AlaProAspAsnLysProAlaProGlySer Thr AlaPro 91 ▶ ProAlaHis GlyVal Thr Ser AlaProAspThr ArgProAlaPro 316 GGTAGTACCGCCCCTCCTGCCCATGGTGTCACATCTGCCCCGGAC 106 FGI vSer Thr AlaProProAlaHis GI vVa I Thr Ser AlaProAsp 361 AACAGGCCTGCATTGGGTAGTACAGCACCGCCAGTACACAACGTT 121 AsnArgProAl aLeuGlySer Thr Al aProProVal Hi sAsnVal 406 ACTACTCCCTCAGGCTCTGCTAGCGGCTCAGCTTCTACTCTGGTG 136 Thr Ser Al aSer Gl vSer Al aSer Gl vSer Al aSer Thr LeuVal 451 CACAACGGCACCTCTGCGCGCGCGACCACAACCCCAGCGAGCAAG 151 Hi sAsnGl yThr Ser Al a ArgAl aThr Thr Thr ProAl aSer Lys 496 AGCACTCCATTCTCAATTCCCAGCTGATAA

166 Ser Thr ProPheSer HeProSer · · · · ·

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Figure 3

1 ATGGGCTCAGCTTCTACTCTGGTGCACAACGGCACCTCTGCCAGG 1 Met Gl vSer Al a Ser Thr LeuVal Hi s Asn Gl vThr Ser Al a Ard 46 GCTACCACAACCCCAGCCAGCAAGAGCACTCCATTCTCAATTCCC 16 AlaThr Thr Thr ProAlaSer LysSer Thr ProPheSer II ePro 91 AGCCACCACTCTGATACTCCTACCACCCTTGCCAGCCATAGCACC 31 Ser Hi sHi s Ser AspThr ProThr Thr Leu Al aSer Hi s Ser Thr 136 AAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCTCCTCTC 46 LysThrAspAl aSer Ser Thr His His Ser Thr Val ProProLeu 181 ACCTCCTCCAATCACAGCACTTCTCCCCAGTTGTCTACTGGGGTC 61 Thr Ser Ser AsnHi s Ser Thr Ser ProGl nLeuSer Thr Gi vVa i 226 TCTTTCTTTTCCTGTCTTTTCACATTTCAAACCTCCAGTTTAAT 76 Ser PhePhePheLeuSer PheHis HeSer AsnLeuGlnPheAsn 271 TCCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAGCTGCAG 91 ▶ Ser Ser LeuGl uAspProSer ThrAspTvrTvrGlnGl uLeuGln 316 AGAGACATTTCTGAAATGTTTTTCCAGATTTATAAACAAGGGGGT 106 ArgAspileSer GluMetPheLeuGinlieTyrLysGinGlyGly 361 TTTCTGGGCCTCTCCAATATTAAGTTCAGGCCAGGATCTGTGGTG 121 PheLeuGivLeuSerAsniieLysPheArgProGivSerVaiVai 406 GTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAATGTCCAC 136 Val GinLeuThr LeuAl aPhe Aro GiuGivThr ille AsnVal His 451 GACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCAGCCTCT 151 AspVa i GluThr GlnPheAsnGlnTvrLvsThr GluAlaAlaSer 496 CGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGATGTGCCA 166 ArgTyrAsnLeuThr HeSerAspValSer ValSerAspValPro 541 TTTCCTTTCTCTGCCCAGTCTGGGGCTGGGGTGCCAGGCTGGGGC 181 ▶ PheProPheSer Al aGI nSer Gi vA I aGI yVa I ProGI v TrpGI v 586 ATCGCGCTGCTGCTGCTCTGTGTTCTGGTTGCGCTGGCCATT 196 FileAlaLeuLeuValLeuValCvsValLeuValAlaLeuAlalle 631 GTCTATCTCATTGCCTTGTGATAA

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Figure 4

271 GCCAACTTGTGATAA 91 AlaAsnLeu••••• Figure 5

1 ATGACAGGTTCTGGTCATGCAAGCTCTACCCCAGGTGGAGAAAAG 1 Met Thr GlySer GlyHisAlaSer Ser Thr ProGlyGlyGluLys 46 GAGACTTCGGCTACCCAGAGAAGTTCAGTGCCCAGCTCTACTGAG 16 FGI uThr Ser Al aThr GI nArg Ser Ser Val ProSer Ser Thr GI u 91 AAGAATGCTGTGAGTATGACCAGCGGCGTACTCTCCAGCCACAGC 31 LvsAsnAl aVal Ser Met Thr Ser Ser Val LeuSer Ser His Ser 136 CCCGGTTCAGGCTCCTCCACCACTCAGGGACAGGATGTCACTCTG 46 ProGlySer GlySer Ser Thr Thr GlnGlyGlnAspVal Thr Leu 181 GCCCGGCCACGGAACCAGCTTCAGCTTCAGCTGCCACCTGGGGA 61 AlaProAlaThr GiuProAlaSer GivSer AlaAlaThr TrpGiv 226 CAGGATGTCACCTCGGTCCCAGTCACCAGGCCAGCCCTGGGCTCC 76 FGI nAspVal Thr Ser Val ProVal Thr ArgProAl aLeuGl ySer 271 ACCACCCGGCAGCCCAGGATGTCACCTCAGCCCCGGACAACAAG 91 Thr Thr ProProAl aHi sAspVal Thr Ser Al aProAspAsnLys 316 CCAGCCCCGGGAAGTACCGCTCCACCACCACCACGCTGTTACCTCG 106 ProAlaProGlySerThrAlaProProAlaHisGlyValThrSer 361 GCTCCGGATACCAGGCCGGCCCCAGGTAGTACCGCCCCTCCTGCC 121 AlaProAspThrArqProAlaProGlySerThrAlaProProAla 406 CATGGTGTCACATCTGCCCCGGACAACAGGCCTGCATTGGGTAGT 136 HisGlyValThr Ser AlaProAspAsnArgProAlaLeuGlySer 451 ACAGCACCGCCAGTACACACGTTACTAGTGCCTCAGGCTCTGCT 151 Thr AlaProProVal His AsnVal Thr Ser AlaSer GlySer Ala 496 AGCGGCTCAGCTTCTACTCTGGTGCACAACGGCACCTCTGCGCGC 166 Fer GlySer AlaSer Thr LeuVal His AsnGlyThr Ser AlaArg 541 GCGACCACACCCCAGCGAGCAAGAGCACTCCATTCTCAATTCCC 181 AlaThr Thr Thr ProAlaSer LysSer Thr ProPheSer I LePro 586 AGCCACCACTCTGATACTCCTACCACCCTTGCCAGCCATAGCACC 196 Fer His His Ser AspThr ProThr Thr LeuAl a Ser His Ser Thr 631 AAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCTCCTCTC 211 LysThrAspAlaSer Ser Thr HisHisSer Thr Val ProProLeu 676 ACCTCCTCCAATCACAGCACTTCTCCCCAGTTGTCTACTGGGGTC 226 Thr Ser Ser AsnHis Ser Thr Ser ProGinLeuSer Thr GlyVal 721 TCTTTCTTTTCCTGTCTTTTCACATTTCAAACCTCCAGTTTAAT 241 ▶ Ser PhePhePheLeuSer PheHis I (eSerAsnLeuGInPheAsn 766 TCCTCTCGGAAGATCCCAGCACCGACTACTACCAAGAGCTGCAG 256 Ser Ser LeuGl uAspProSer Thr AspTyrTyrGl nGl uLeuGl n 811 AGAGACATTTCTGAAATGTTTTTGCAGATTTATAAACAAGGGGGT 271 ArgAsplieSer GluMetPheLeuGinlieTyrLysGlnGlyGly 856 TTTCTGGGCCTCTCCAATATTAAGTTCAGGCCAGGATCTGTGGTG 286 PheLeuGlyLeuSerAsnlleLysPheArgProGlySerValVal

(Continued)

Figure 5 (continued)

901 GTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAATGTCCAC 301 Val GinLeuThr LeuAl aPhe ArgGiuGiyThr HeAsnVal His 946 GACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCAGCCTCT 316 AspVal GluThr GlnPheAsnGlnTyrLysThr GluAlaAlaSer 991 CGATATAACCTGACGATCTCAGACGTCAGCGTGAGTGATGTGCCA 331 ArgTyrAsnLeuThr HeSerAspValSerValSerAspValPro 1036 TTTCCTTTCTCTCCCCAGTCTGGGGCTGCGGGTGCCAGGCTGGGGC 346≯PheProPheSer Al aGl nSer Gl yAl aGl yVal ProGl yTrpGl y 1081 ATCGCGCTGCTGGTGCTGGTCTGTGTTCTGGTTGCGCTGGCCATT 1126 GTCTATCTCATTGCCTTGGCTGTCTGTCAGTGCCGCCGAAAGAAC 376 Val TyrLeu i leAl aLeuAl aVal CysGlnCysArg ArgLysAsn 391 TyrGiyGinLeuAspliePheProAlaArgAspThrTyrHisPro 1216 ATGAGCGAGTACCCCACCTACCACCCCATGGGCGCTATGTGCCC 406 Met Ser GluTyrProThr TyrHisThr HisGlyArgTyrVal Pro 1261 CCTAGCAGTACCGATCGTAGCCCCTATGAGAAGGTTTCTGCAGGT 421 ProSer Ser ThrAspArgSer ProTyrGluLysVal Ser AlaGiv 1306 AATGGTGGCAGCAGCCTCTCTTACACAAACCCAGCAGTGGCAGCC 436 AsnGl vGl vSer Ser LeuSer TyrThr AsnProAlaValAlaAla 1351 ACTTCTGCCAACTTGTGATAA 451 Thr Ser Al aAsnLeu•••••

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Figure 6

361 CGAGGATCC 121 A rgGi ySer

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC 1 Met Giniie Phe Val Lys Thr Leu Thr Gly Lys Thr He Thr Leu 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC 16 ▶ GluVal GluProSerAspThrileGluAsnValLysAlaLysile 91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT 31 ▶ GIn AspLysGiuGly IIeProProAspGinGin ArgLeu IIePhe 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC 46 A laGlyLysGlnLeuGluAspGlyArgThrLeuSerAspTyrAsn 181 ATCCAGAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT 61. I leGinLvsGluSer Thr LeuHisLeuVal LeuArgLeuArgGly 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTG 76 ➤ GlyArgHisGlySerGlyAlaTrpLeuLeuProValSerLeuVal 271 AAAAGAAAACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCC 91 ▶ Lys ArgLysThr Thr LeuAl aProAsnThr GlnThr Al aSer Pro 316 CGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCC 106 A rgAl aLeuAl aAspSer LeuMet Gl nLeuAl aArgGl nVal Ser

Figure 7

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC 1 Met Ginile Phe Vai Lys Thr Leu Thr Gly Lys Thr He Thr Leu 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC 16 GluValGluProSerAspThrlleGiuAsnValLvsAlaLvsile 91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT 31 GinAspLysGiuGiylieProProAspGinGinArgLeuilePhe 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC 46 A I aGi yLysGi nLeuGi uAspGi yA rgThr LeuSerAspTyrAsn 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT 61 ► I I eGI nLysGI uSer Thr LeuHi sLeuVal LeuArgLeuArgGI v 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTG 76 GlyArgHisGlySer GlyAlaTrpLeuLeuProValSer LeuVal 271 AAAAGAAAAACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCC 91 Lys ArgLysThr Thr LeuAi aProAsnThr GinThr AlaSer Pro 31.6 CGCCCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCC 106 A rgAl aLeuAl aAspSer LeuMet GlinLeuAl aArgGlinVal Ser 361 CGAGGATCCACAGGTTCTGGTCATGCAAGCTCTACCCCAGGTGGA 121 A rgGl ySer Thr Gl ySer Gl yHi s Al aSer Ser Thr ProGl yGl y 406 GAAAAGGAGACTTCGGCTACCCAGAGAAGTTCAGTGCCCAGCTCT 136 FGI uLvsGI uThr Ser Al aThr GI n ArgSer Ser Val ProSer Ser 451 ACTGAGAAGAATGCTGTGAGTATGACCAGCAGCGTACTCTCCAGC 151 Thr GluLvs AsnAl aVal Ser Met Thr Ser Ser Val LeuSer Ser 496 CACAGCCCCGGTTCAGGCTCCTCCACCACTCAGGGACAGGATGTC 166 His Ser ProGlySer GlySer Ser Thr Thr GlnGlyGlnAspVal 541 ACTCTGGCCCGGCCACGGAACCAGCTTCAGGTTGATAA 181 Thr LeuAl aProAl aThr GluProAl aSer Gly *****

Figure 8

-1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC 1 Met GinilePheValLysThr LeuThr GiyLysThr | leThr Leu 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC 16 GluVal GluProSerAspThr HeGluAsnValLysAlaLysHe 91 CAAGACAAGGAAGCATCCCTCCTGACCAGCAGAGGCTCATCTTT 31 GI nAspLysGI uGI y I I eProProAspGI nGI nArgLeu I I ePhe 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC 46 A I aGI vLvsGI nLeuGI uAspGI vA rgThr LeuSerAspTvrAsn 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT 61 I i eGi nLvsGi uSer Thr LeuHi sLeuVa i LeuAroLeuAroGi v 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTG 76 GIVA rgHis GIVSer GIVA I aT rpLeuLeuProVal Ser LeuVai 271 AAAAGAAAAACCACCCTGGCGCCCCAATACGCAAACCGCCTCTCCC 91 Lys ArgLysThr Thr Leu Al a Pro AsnThr GlnThr Al a Ser Pro 316 CGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCC 106 A rgAl aLeuAl aAspSer LeuMet GinLeuAl aArgGinVal Ser 361 CGAGGATCCGTGCCCAGCTCTACTGAGAAGAATGCTGTGAGTATG 121 A roGI vSer Val ProSer Ser Thr GI uLvsAsnAl aVal Ser Met 406 ACCAGCAGCGTACTCTCCAGCCACAGCCCCGGTTCAGGCTCCTCC 136 Thr Ser Ser Val LeuSer Ser Hils Ser ProGlySer GlySer Ser 451 ACCACTCAGGGACAGGATGTCACTCTGGCCCCGGCCACGGAACCA 151 Thr Thr GinGiyGinAspVaiThr LeuAiaProAlaThr GiuPro 496 GCTTCAGGTTCAGCTGCCACCTGGGGACAGGATGTCACCTCGGTC 166 AlaSer GlySer AlaAlaThr TrpGlyGlnAspValThr Ser Val 541 CCAGTCACCAGGCCAGCCCTGGGCTCCACCACCCCGCCAGCCCAC 181 ProVai Thr ArgProAl aLeuGi ySer Thr Thr ProProAl aHi s 586 GATGTCACCTCAGCCCCGGACAACAAGCCAGCCCCGGGAAGTACT 196 AspVa | Thr Ser Al aProAspAsnLysProAl aProGlySer Thr 631 GCTCCACCAGCACACGGTGTTACCTCGGCTCCGGATACCAGGCCG 211 AlaProProAlaHisGiyValThr Ser AlaProAspThr ArgPro 676 GCCCCAGGTAGTACCGCCCCTCCTGCCCATGGTGTCACATCTGCC 226 AlaProGlySer Thr AlaProProAlaHis GlyVal Thr Ser Ala 721 CCGGACAACAGGCCTGCATTGGGTAGTACAGCACCGCCAGTACAC 241 ProAspAsnArgProAlaLeuGlySerThr AlaProProValHis 766 AACGTTACTAGTGCCTCAGGCTCTGCTAGCGGCTCAGCTTCTACT 256 AsnVal Thr Ser AlaSer GlySer AlaSer GlySer AlaSer Thr 811 CTGGTGCACAACGCACCTCTGCGCGCGCGACCACAACCCCAGCG 271 LeuVal Hi sasnGl vThr Ser Ala Argal aThr Thr Thr ProAla 856 AGCAAGAGCACTCCATTCTCAATTCCCAGCTGATAA 286 Ser LysSer Thr ProPheSer | LeProSer

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Figure 9

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC 1 MetGinilePheValLysThrLeuThrGiyLysThrlieThrLeu 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC 16 PGI uVal GI uProSerAspThrileGi uAsnVal LvsAl aLvsile 91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT 31 GinAspLysGiuGiyIieProProAspGinGinArgLeuliePhe 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC 46 A I aGI yLysGI nLeuGI uAspGI yA rgThr LeuSerAspTyrAsn 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT 61 ► I i eGi nLysGl uSer Thr LeuHi sLeuVal LeuArgLeuArgGl y 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGTG 76 FGI yArgHis GI ySer GI yAl aTrpLeuLeuProVal Ser LeuVal 271 AAAAGAAAAACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCC 91▶LysArgLysThr Thr LeuAlaProAsnThr GinThr AlaSer Pro 316 CGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCC 106▶ ArgAlaLeuAlaAspSerLeuMetGinLeuAlaArgGinValSer 361 CGAGGATCCGGCTCAGCTTCTACTCTGGTGCACAACGGCACCTCT 121 ArgGlySer GlySer AlaSer Thr LeuVal HisAsnGlyThr Ser 406 GCCAGGGCTACCACAACCCCAGCCAGCAAGAGCACTCCATTCTCA 136 AlaArgAlaThr Thr Thr ProAlaSer LysSer Thr ProPheSer 151 ▶ | LeProSer HisHis Ser AspThr ProThr Thr LeuAlaSer His 496 AGCACCAAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCT 166▶ Ser Thr LysThrAspAl aSer Ser Thr Hi sHl s Ser Thr Va! Pro 541 CCTCTCACCTCCTACCACCACCACCTCCCCCAGTTGTCTACT 181 ProLeuThr Ser Ser AsnHi s Ser Thr Ser ProGinLeuSer Thr 586 GGGGTCTCTTTCTTTTTCCTGTCTTTTCACATTTCAAACCTCCAG 196▶ GlyVal Ser PhePhePheLeuSer PheHls i LeSer AsnLeuGl n 631 TTTAATTCCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAG 211▶ PheAsnSer Ser LeuGl uAspProSer Thr AspTyrTyrGl nGl u 676 CTGCAGAGAGACATTTCTGAAATGTTTTTGCAGATTTATAAACAA 226 ▶ LeuGin ArgAspileSer GluMet PheLeuGin Ite TyrLysGln 721 GGGGGTTTTCTGGGCCTCTCCAATATTAAGTTCAGGCCAGGATCT 241 FGI yGI yPheLeuGI yLeuSerAsn! I eLysPheArgProGI ySer 766 GTGGTGGTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAAT 256▶ Val Val Val GinLeuThr LeuAl aPhe ArgGiuGiyThr IteAsn 811 GTCCACGACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCA 271 ▶ Val Hi sAspVal Gl uThr Gl nPheAsnGl nT yrLysThr Gl uAl a 286▶AlaSerArgTyrAsnLeuThrlleSerAspValSerValSerAsp 901 GTGCCATTTCTTTCTCTGCCCAGTCTGGGGCTGGGGTGCCAGGC 301 Val ProPheProPheSer AlaGinSer GiyAlaGiyVal ProGiy 946 TGGGGCATCGCGCTGCTGGTGCTGTTCTGGTTGCGCTG 316 TrpGiyileAlaLeuLeuValLeuValCysValLeuValAlaLeu 991 GCCATTGTCTATCTCATTGCCTTGTGATAA 331 AlalieVaiTyrLeulleAlaLeu....

Figure 10

1 ATGCAGATCTTCGTGAAGACCCTGACTGGTAAGACCATCACTCTC 1 MetGInItePheValLvsThr LeuThr GlvLvsThr HeThr Leu 46 GAAGTGGAGCCGAGTGACACCATTGAGAATGTCAAGGCAAAGATC 16 GiuVai GiuProSerAspThrileGiuAsnVaiLysAlaLyslie 91 CAAGACAAGGAAGGCATCCCTCCTGACCAGCAGAGGCTCATCTTT 31 ▶ Gi nAspLvsGi uGi vi i eProProAspGi nGi nArqLeu i i ePhe 136 GCAGGCAAGCAGCTGGAAGATGGCCGCACTCTTTCTGACTACAAC 46 AlaGiyLysGinLeuGiuAspGiyArgThrLeuSerAspTyrAsn 181 ATCCAGAAAGAGTCCACCCTGCACCTGGTGCTCCGTCTCAGAGGT 61 ► I I eGI nLvsGI uSer Thr LeuHi sLeuVa I LeuArgLeuArgGI v 226 GGGAGGCACGGTAGTGGTGCATGGCTGTTGCCCGTCTCGCTGGTG 76 Gi yArgHi s Gi ySer Gi yAl aTrpLeuLeuProVal Ser LeuVal 271 AAAAGAAAAACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCC 91 Lys ArgLysThr Thr Leu Al a Pro AsnThr GinThr Al a Ser Pro 316 CGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCC 106 A roAl aLeuAl aAspSer LeuMet GlinLeuAl aAroGinVal Ser 361 CGAGGATCCCTGGTGCTGGTCTGTGTTCTGGTTGCGCTGGCCATT 121 A rgGiySer LeuVal LeuVal CysVal LeuVal Al aLeuAl al le 406 GTCTATCTCATTGCCTTGGCTGTCTGTCAGTGCCGCCGAAAGAAC 136 VaiTyrLeuileAlaLeuAlaValCysGinCysArgArgLysAsn 451 TACGGCAGCTGGACATCTTTCCAGCCCGGGATACCTACCATCCT 151 TvrGlvGlnLeuAspllePheProAlaArgAspThrTvrHisPro 496 ATGAGCGAGTACCCCACCTACCACACCCATGGGCGCTATGTGCCC 166 Met Ser GluTvrProThr TvrHis Thr His GlvArqTvrVa (Pro 541 CCTAGCAGTACCGATCGTAGCCCCTATGAGAAGGTTTCTGCAGGT 181 ProSer Ser Thr Asp Arg Ser ProTyr GluLys Val Ser Ala Gly 586 AATGGTGCAGCAGCCTCTCTTACACAAACCCAGCAGTGGCAGCC 196 AsnGlyGlySer Ser LeuSer TyrThr AsnProAlaVal AlaAla 631 ACTTCTGCCAACTTGTGATAA 211 Thr Ser Al aAsnLeu • • • • •

Figure 11

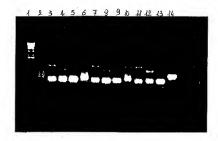
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Figure 11 (continued)

901 GCGCGCGCGACCACAACCCCAGCGAGCAAGAGCACTCCATTCTCA 301 AlaArgAlaThr Thr Thr ProAlaSer LysSer Thr ProPheSer 316 I Je Pro Ser Hi s Hi s Ser Asp Thr Pro Thr Thr Leu Al a Ser Hi s 991 ACCACCAAGACTGATGCCAGTAGCACTCACCATAGCACGGTACCT 331▶ Ser Thr LysThrAspAl aSer Ser Thr HisHis Ser Thr Val Pro 1036 CCTCTCACCTCCTCCAATCACAGCACTTCTCCCCAGTTGTCTACT 346 ProLeuThr Ser Ser AsnHis Ser Thr Ser ProGin LeuSer Thr 1081 GGGGTCTCTTTCTTTTTCCTGTCTTTTCACATTTCAAACCTCCAG 361 FGI vVa i Ser PhePhePheLeuSer PheHi s I I eSer AsnLeuGI n 1126 TTTAATTCCTCTCTGGAAGATCCCAGCACCGACTACTACCAAGAG 376) PheAsnSer Ser LeuGl uAspProSer ThrAspTyrTyrGl nGl u 1171 CTGCAGAGAGACATTTCTGAAATGTTTTTGCAGATTTATAAACAA 391 ▶ LeuGin ArgAspiieSer GluMet PheLeuGiniieTyrLysGin 1216 GGGGGTTTTCTGGGCCTCTCCAATATTAAGTTCAGGCCAGGATCT 406 FGI yGI yPheLeuGI yLeuSerAsnII eLysPheArgProGI ySer 1261 GTGGTGGTACAATTGACTCTGGCCTTCCGAGAAGGTACCATCAAT 421 Val Val Val GinLeuThr LeuAl aPhe ArgGiuGiyThr ileAsn 1306 GTCCACGACGTGGAGACACAGTTCAATCAGTATAAAACGGAAGCA 436 Val Hi saspVal GluThr GinPheAsnGlnTyrLysThr GluAl a 451 AlaSer ArgTyrAsnLeuThr IleSerAspValSer ValSerAsp 1396 GTGCCATTTCCTTTCTCTGCCCAGTCTGGGGCTGGGGTGCCAGGC 466 Val ProPheProPheSer Al aGl nSer Gl yAl aGl yVal ProGl y 1441 TGGGGCATCGCGCTGCTGGTGCTGTGTTCTGGTTGCGCTG 481 TrpGivIIeAlaLeuLeuValLeuValCysValLeuValAlaLeu 1486 GCCATTGTCTATCTCATTGCCTTGGCTGTCTGTCAGTGCCGCCGA 496 Al'alle Val TyrLeulle Al aLeu Al a Val Cys Gln Cys Arg Arg 1531 AAGAACTACGGGCAGCTGGACATCTTTCCAGCCCGGGATACCTAC 511 LvsAsnTvrGlvGlnLeuAspllePheProAlaArgAspThrTyr 1576 CATCCTATGAGCGAGTACCCCACCTACCACACCCATGGGCGCTAT 526 His ProMet Ser GluTyrProThr TyrHis Thr His GlyArgTyr 1621 GTGCCCCCTAGCAGTACCGATCGTAGCCCCTATGAGAAGGTTTCT 541 ▶ Val ProProSer Ser ThrAspArgSer ProTyrGluLysVal Ser 1666 GCAGGTAATGGTGGCAGCAGCCTCTCTTACACAAACCCAGCAGTG 556 AlaGiyAsnGiyGiySer Ser LeuSer TyrThrAsnProAlaVal 1711 GCAGCCACTTCTGCCAACTTGTGATAA

571 Al aAl aThr Ser Al aAshLeu

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Figure 13

1	CCAGGAAGCTCCTCTGTGTCCTCATAAACCCTAACCTCCTCTACTTGAG
51	GGACATTCCAATCATAGGCTGCCCATCCACCCTCTGTGTCCTCCTGTTA
101	TTAGGTCACTTAACAAAAAGGAAATTGGGTAGGGGTTTTTCACAGACCG
151	TTTCTAAGGGTAATTTTAAAATATCTGGGAAGTCCCTTCCACTGCTGTG
201	TCCAGAAGTGTTGGTAAACAGCCCACAAATGTCAACAGCAGAAACATAC
251	AGCTGTCAGCTTTGCACAAGGGCCCAACACCCTGCTCATCAAGAAGCAC
301	GTGGTTGCTGTTAGTAATGTGCAAAACAGGAGGCACATTTTCCCCAC
351	TGTGTAGGTTCCAAAATATCTAGTGTTTTCATTTTTACTTGGATCAGGA
401	CCCAGCACTCCACTGGATAAGCATTATCCTTATCCAAAACAGCCTTGTG
451	${\tt TCAGTGTTCATCTGCTGACTGTCAACTGTAGCATTTTTTGGGGTTACAGGGATAGGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGG$
501	TTGAGCAGGATATTTGGTCCTGTAGTTTGCTAACACACCCTGCAGCTCC
551	AAGGTTCCCCACCAACAGCAAAAAAATGAAAATTTGACCCTTGAATGGG
601	TTTCCAGCACCATTTTCATGAGTTTTTTGTGTCCCTGAATGCAAGTTTA
651	CATAGCAGTTACCCCAATAACCTCAGTTTTAACAGTAACAGCTTCCCAC
701	${\tt TCAAAATATTTCCACAGGTTAAGTCCTCATTTAAATTAGGCAAAGGAATTAGGAAAGGAATTAGGAAAGGAATTAGGAAAGGAATTAGGAAAGGAATTAGGAAAGGAATTAGGAAAGGAATTAGGAAAGGAAAGGAATTAGGAAAGGAAAGGAAAGGAATTAGGAAAGGAAAGGAAAGGAAATTAGGAAAGGAAATTAGGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAA$
751.	$\tt CTTGAAGACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCTCATTTTTATAGGTTAATGTCTCATTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTTATAGGTTAATGTCTCATTTTTTTT$
801	${\tt ATGATAATGGTTTCTTAGACGTCAGGTGGCACTTTTCGGGGAAATGGTGGGGAAATGGTGGGGGGGG$
851	GCGCGGAACCCCTATTTGTTTATTTTTCTAAATACATTCAAATATGTATC
901	CGCTCATGAGACAATAACCCTGATAAATGCTTCAATAATATTGAAAAAGC
951	${\tt AAGAGTATGAGTATTCAACATTTCCGTGTCGCCCTTATTCCCTTTTTTGCCTTTTTTTGCCTTTTTTTGCCTTTTTT$
1001	GGCATTTTGCCTCCTGTTTTTGCTCACCCAGAAACGCTGGTGAAAGTAA

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Figure 13

2151 TAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTACATACCTCGC .2201 TCTGCTAATCCTGTTACCAGTGGCTGCCAGTGGCGATAAGTCGTGTC 2251 TTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTCG 2301 GGCTGAACGGGGGTTCGTGCACACAGCCCAGCTTGGAGCGAACGACCTA 2351. CACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTC 2401 CCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTCGGAACA 2451 GGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCCTGGTATCTTTATAG 2501 TCCTGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGCT 2551 CGTCAGGGGGGGGGGGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTA 2601 CGGTTCCTGGCCTTTTGCTGGCCTTTTGCTCACATGFTCTTTCCTGCGTT 2701 CCGCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAA 2751 GCGGAAGAGCGCCTGATGCGGTATTTTCTCCTTACGCATCTGTGCGGTAT 2801 TTCACACOGCATATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGCAT 2851 AGTTAAGCCAGTATACAATCAATATTGGCCATTAGCCATATTATTCATTG 2901 GTTATATAGCATAAATCAATATTGGCTATTGGCCATTGCATACGTTGTAT 2951 CCATATCATAATATGTACATTTATATTGGCTCATGTCCAACATTACCGCC 3001 ATGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGT 3051 CATTAGTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTA 3101 AATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCCATTGACGTCAAT 3151 AATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTC 3201 AATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTG 3251 TATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCC (Continued)

Figure 13 (Continued)

3301 CGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGC 3351 AGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGCGGTTTTGG 3401 CAGTACATCAATGGGCGTGGATAGCGGTTTGACTCACGGGGATTTCCAAG 3451 TCTCCACCCCATTGACGTCAATGGGAGTTTGTTTTGGCACCAAAATCAAC 3501 GGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAAATGGGC 3551 GCTAGGCGTGTACGCTGGGAGGTCTATATAAGCAGAGCTCGTTTAGTGAA 3601 CCGTCAGATCGCCTGGAGACGCCATCCACGCTGTTTTGACCTCCATAGAA 3651 GACACCGGGACCGATCCAGCCTCCGCGGCCGGGAACGGTGCATTGGAACG 3701 CGGATTCCCCGTGCCAAGAAGCTTGTCTAGAACCCGGGAGAGCTCCTGA 3751 GAACTTCAGGGTGAGTTTGGGGACCCTTGATTGTTCTTTTTTTCGCTA 3801 TTGTAAAATTCATGTATATGGAGGGGGGAAAGTTTTCAGGGTGTTTTTT 3851 AGAATGGGAAGATGTCCCTTGTATCACCATGGACCCTCATGATAATTTTG 3901 TTTCTTTCACTTTCTACTCTGTTGACAACCATTGTCTCCTCTTATTTTCT 3951 TTTCATTTTCTGTAACTTTTCGTTAAACTTTAGCTTGCATTTGTAACGA 4001 ATTTTTAAATTCACTTTTGTTTATTTGTCAGATTGTAAGTACTTTCTCTA 4051 ATCACTTTTTTTCAAGGCAATCAGGGTATATTATATTGTACTTCAGCAC 4101 AGTTTTAGAGAACAATTGTTATAATTAAATGATAAGGTAGAATATTTCTG 4151 CATATAAATTCTGGCTGGCGTGGAAATATTCTTATTGGTAGAAACAACTA 4201 CATCCTGGTCATCATCCTGCCTTTCTCTTTATGGTTACAATGATATACAC 4251 TGTTTGAGATGAGGATAAAATACTCTGAGTCCAAACCGGGCCCCTCTGCT 4301 AACCATGTTCATGCCTTCTTCTTTTTCCTACAGCTCCTGGGCAACGTGCT 4351 GGTTGTTGTGCTGTCTCATCATTTTGGCAAAGAATTCACTCCTCAGGTGC 4401 AGGCTGCCTATCAGAAGGTGGTGGCTGTGTGGCCAATGCCCTGGCTCAC (Continued) The second secon

Figure 13 (Continued)

1051 AAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAACTGGAT 1101 CTCAACAGCGGTAAGATCCTTGAGAGTTTTCGCCCCGAAGAACGTTTTCC 1151 AATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATTATCCCGTG 1201 TTGACGCCGGGCAAGAGCAACTCGGTCGCCGCATACACTATTCTCAGAAT 1.251 GACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCTTACGGATGGCAT 1301 GACAGTAAGAGAATTATGCAGTGCTGCCATAACCATGAGTGATAACACTG 1351 CGGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCT 1401 TTTTTGCACAACATGGGGGATCATGTAACTCGCCTTGATCGTTGGGAACC 1451 GGAGCTGAATGAAGCCATACCAAACGACGAGCGTGACACCACGATGCCTG 1551 CTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGC 1601 AGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATA 1.651 AATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGG 1701 CCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCA 1751 GGCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTGCCTCAC 1801 TGATTAAGCATTGGTAACTGTCAGACCAAGTTTACTCATATATACTTTAG 1851 ATTGATTTAAAACTTCATTTTTAATTTAAAAGGATCTAGGTGAAGATCCT 1901 TTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACT 1951 GAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTT 2001 TTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAAACCACCGCTACCAGC 2051 GGTGGTTTGTTTGCCGGATCAACAGCTACCAACTCTTTTTCCGAAGGTAA 2101 CTGGCTTCAGCAGAGCGCAGATACCAAATACTGTCCTTCTAGTGTAGCCG (Continued)

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Figure 13 (Continued)

4451 AAATACCACTGAGATCTTTTTCCCTCTGCCAAAAATTATGGGGACATCAT
4501 GAAGCCCCTTGAGCATCTGACTTCTGGCTAATAAAGGAAATTATTTTTCA
4551 TTGCAATAGTGTGTTGGAATTTTTTTGTGTCCTCACTCGGAAGGACATAT
4601 GGGAGGGCAAATCATTTAAAACATCAGGATGAGTATTTGGTTTAGAGTTT
4651 GGCAACATATGCCATAGACCACGAACAAAGGTGGCTATAAAGA
4701 GGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTCCTTATTCCATAG
4751 AAAGCCTTGACTTGAGGTTAGATTTTTTTTATTTTTGTTTTGTTTTGTTAG
4801 TTTTTCTTTAACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGA
4851 TTTTTCCTCCTCTCCTGACTACCCCAGTCATAGCTGTCCCTCTCTCC
4901 GATCC

WO 00/25827

SEQUENCE LISTING

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